ELECTROCHEMICAL BIOSENSORS

Modern and future approaches to medical diagnostics

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Medical Diagnostics

- **Doctors increasingly rely on testing**
- **Needs: rapid, cheap, and “low tech”**
- **Done by technicians or patients**
- **Some needs for *in-vivo* operation, with feedback**

Principle of Electrochemical Biosensors

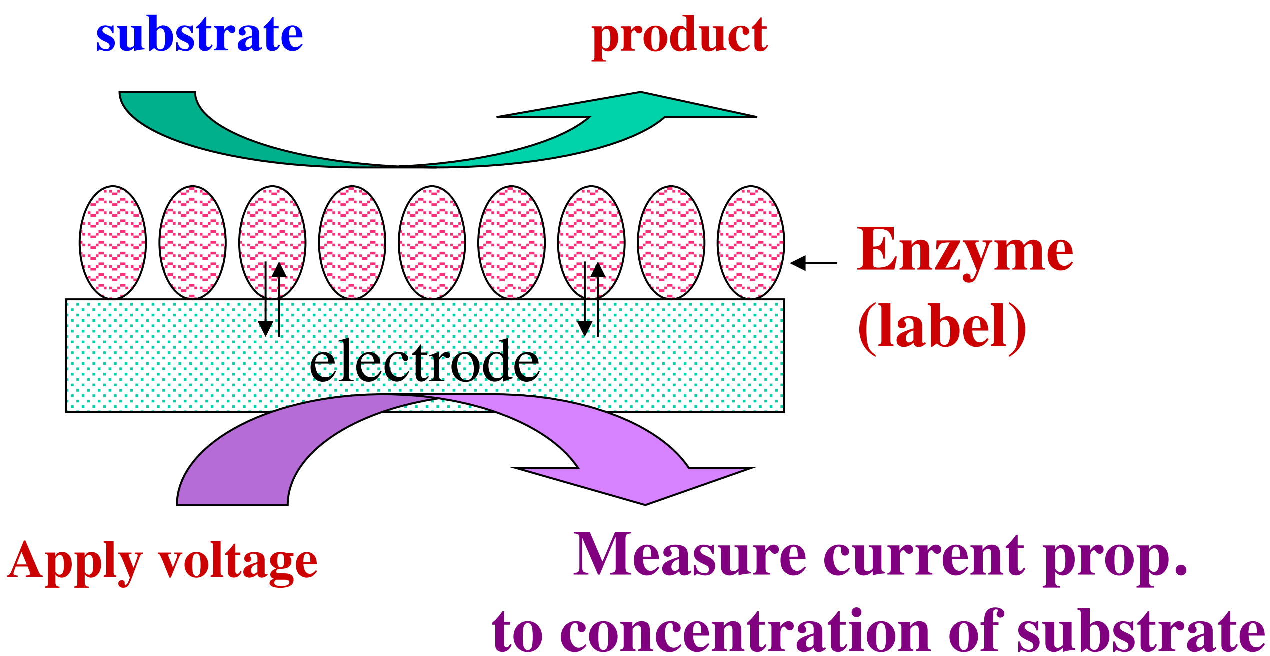
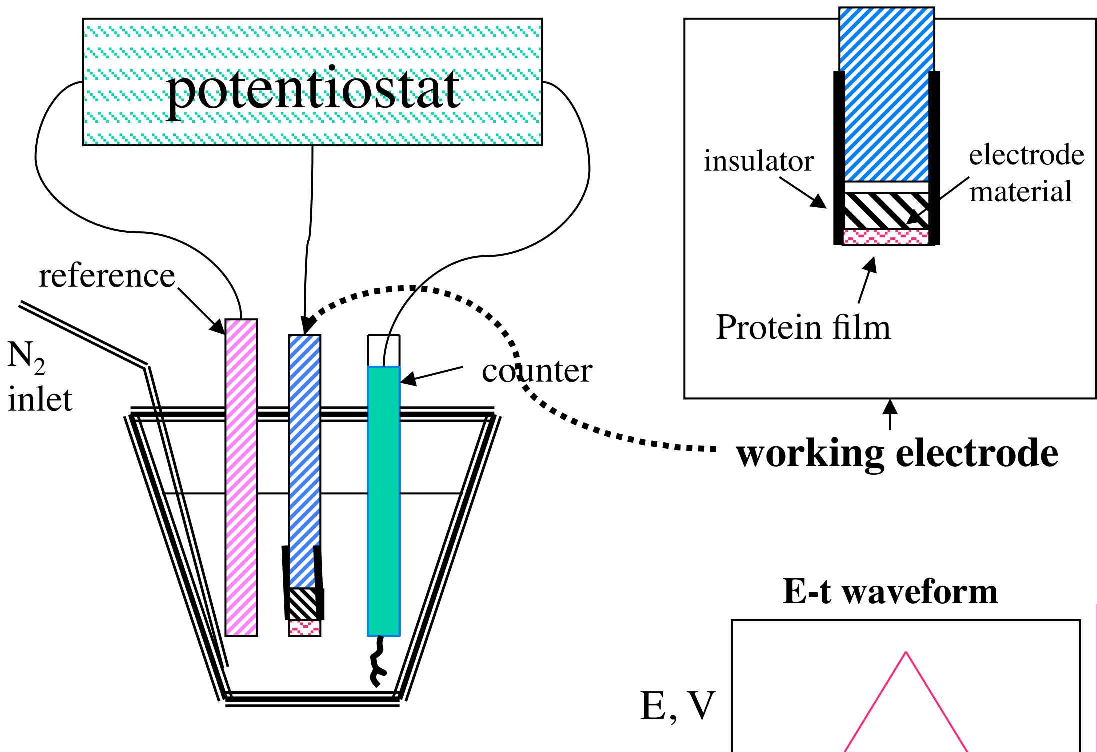
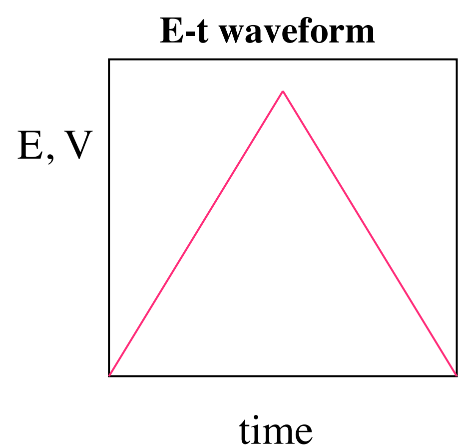


Figure 9

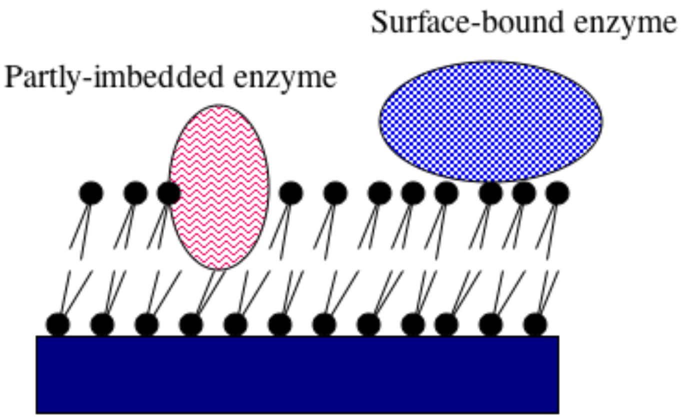
Equipment for developing electrochemical biosensors

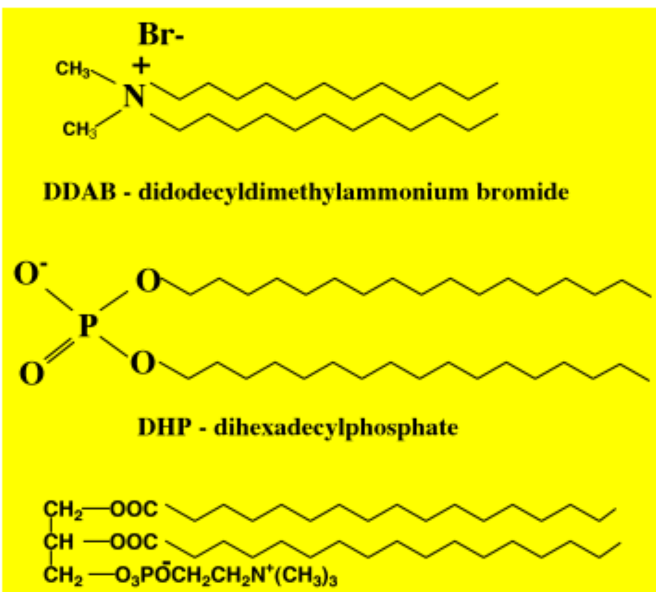


Electrochemical cell



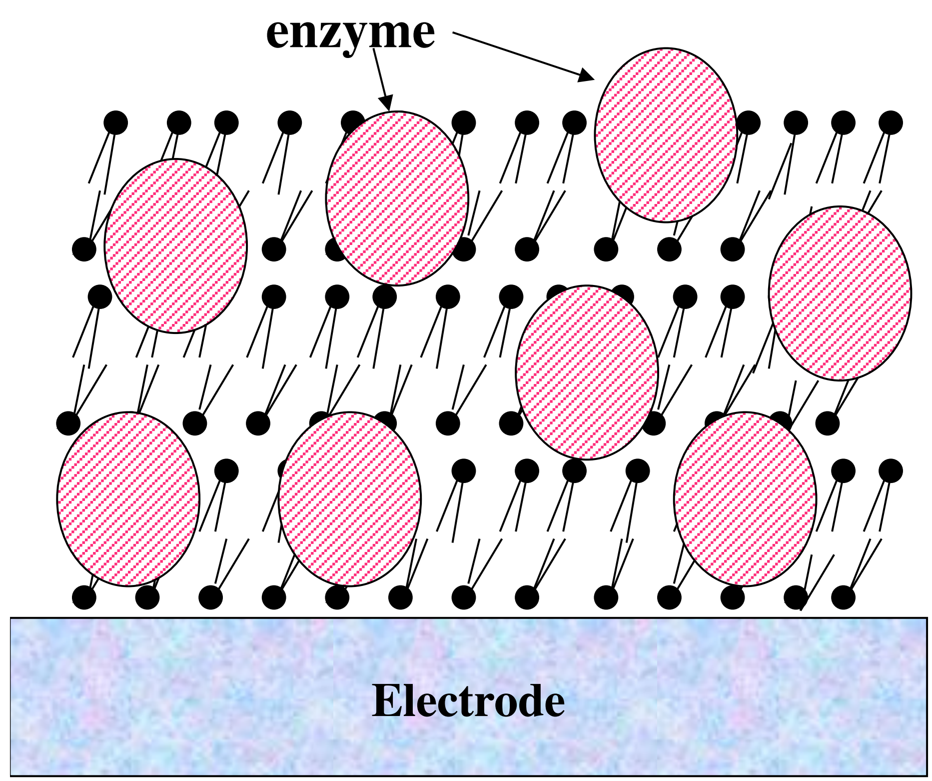
Cyclic voltammetry





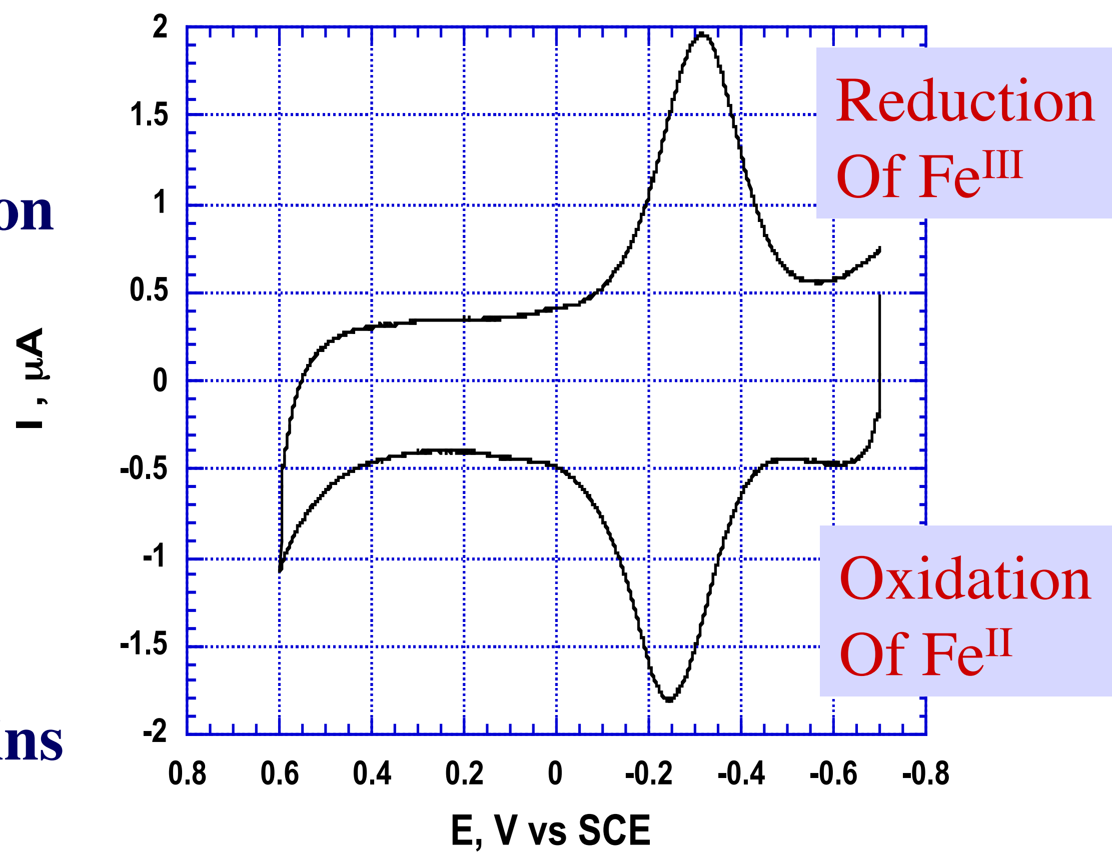
Dihexadecylphosphatidyl choline

A lipid-enzyme film



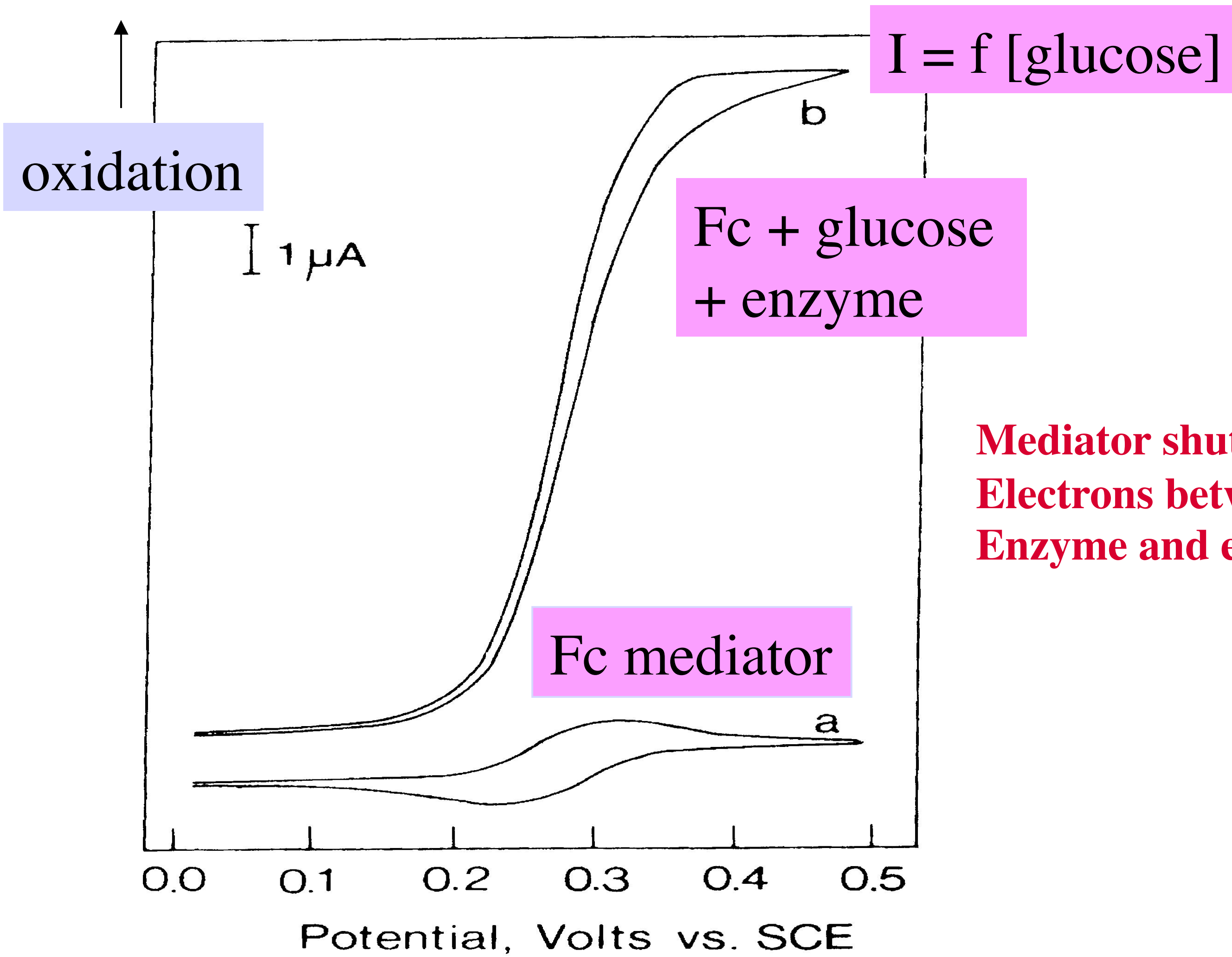
Cyclic voltammogram (CV) at 100 mV s^{-1} and 25 °C of *Mycobacterium Tuberculosis* KatG catalase-peroxidase in a thin film of dimyristoylphosphatidylcholine on basal plane PG electrode, in anaerobic pH 6.0 buffer.

Reversible Peaks for Direct electron Transfer



(not all proteins do this)

Catalytic enzyme electrochemistry
a basis for biosensor - glucose oxidase



Mediator shuttles
Electrons between
Enzyme and electrode

A. Cass, G. Davis, G. D. Francis, H. O. A. Hill, W. J. Aston, I. J. Higgins, E. V. Plotkin, L. D. L. Scott, A. P. F. Turner, *Anal. Chem.* **56,** 667-671 (1984).

Mechanism for catalytic oxidation of glucose With Glucose oxidase (GO) and Fc mediator

Scheme 2

$$\text{Glucose} + \text{GO(FAD)} + 2\,H^+ \rightarrow \text{gluconolactone} + \text{GO(FADH}_2\text{)} \qquad (1)$$

$$\text{GO(FADH}_2\text{)} + 2\,Fc^+ \rightarrow \text{GO(FAD)} + 2\,Fc + 2\,H^+ \qquad (4)$$

$$Fc \rightarrow Fc^+ + 2\,e^- \quad \text{(at electrode)} \quad (5)$$

Fc = ferrocenecarboxylate

Signal can also be measured by amperometry:
Hold const. E where oxidation occurs, measure I vs time

Commercial Glucose Sensors

- **Biggest biosensor success story!**
- **Diabetic patients monitor blood glucose at home**
- **First made by Medisense (early 1990s), now 5 or more commercial test systems**
- **Rapid analysis from single drop of blood**
- **Enzyme-electrochemical device on a slide**

- **Most sensors use enzyme called glucose oxidase (GO)**
- **Most sensors are constructed on electrodes, and use a <u>mediator</u> to carry electrons from enzyme to GO**

Fc = mediator, ferrocene, an iron complex

These reactions occur in the sensor:

$$Fc \longleftrightarrow Fc^{+} + e^{-} \text{ (measured)}$$

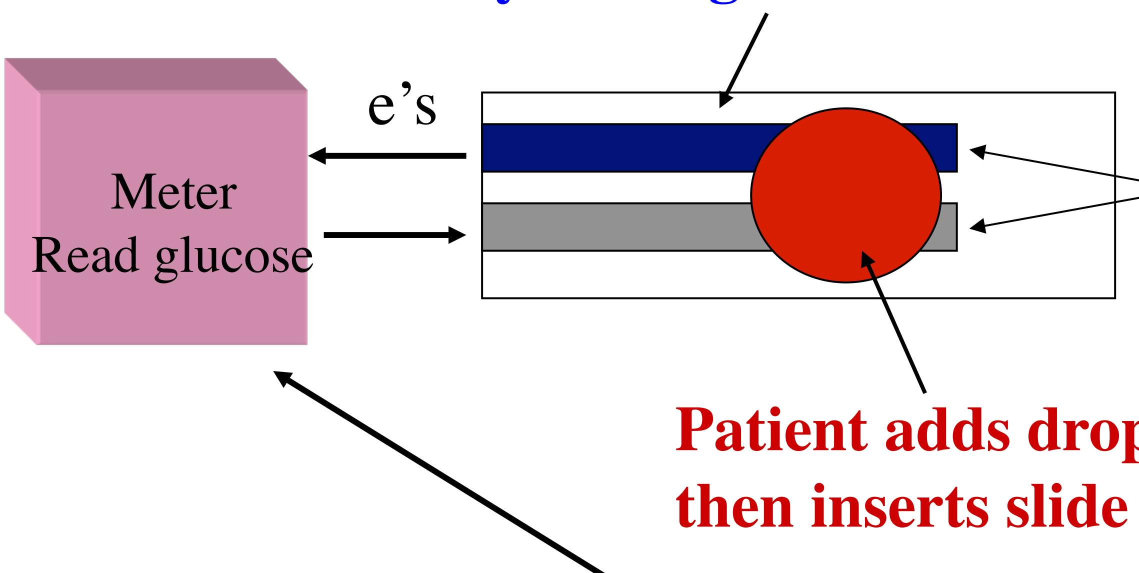
$$GO_{R} + 2\,Fc^{+} \rightarrow GO_{ox} + 2\,Fc$$

$$GO_{ox} + \text{glucose} \rightarrow GO_{R} + \text{gluconolactone}$$

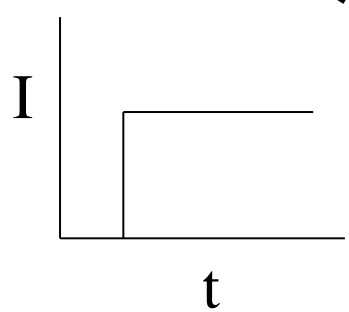
Reach and Wilson, *Anal. Chem.* 64, 381A (1992)

G. Ramsay, *Commercial Biosensors*, J. Wiley, 1998.

Glucose biosensor test strips (~$0.50-1.00 ea.)



Output:
Amperometry
Constant E



Research on glucose sensors

- **Non-invasive biosensors - skin, saliva**
- **Implantable glucose sensors to accompany artificial pancreas - feedback control of insulin supply**
- **Typical use 3-4 weeks for implantable sensor in humans**
- **Failure involved fouling and inflammation**

Other biosensors

- **Cholesterol - based on cholesterol oxidase**
- **Antigen-antibody sensors - toxic substances, pathogenic bacteria**
- **Small molecules and ions in living things: H^+, K^+, Na^+, CO_2, H_2O_2**
- **DNA hybridization and damage**
- **Micro or nanoarrays, optical abs. or fluorescence**

Layer by layer Film construction:

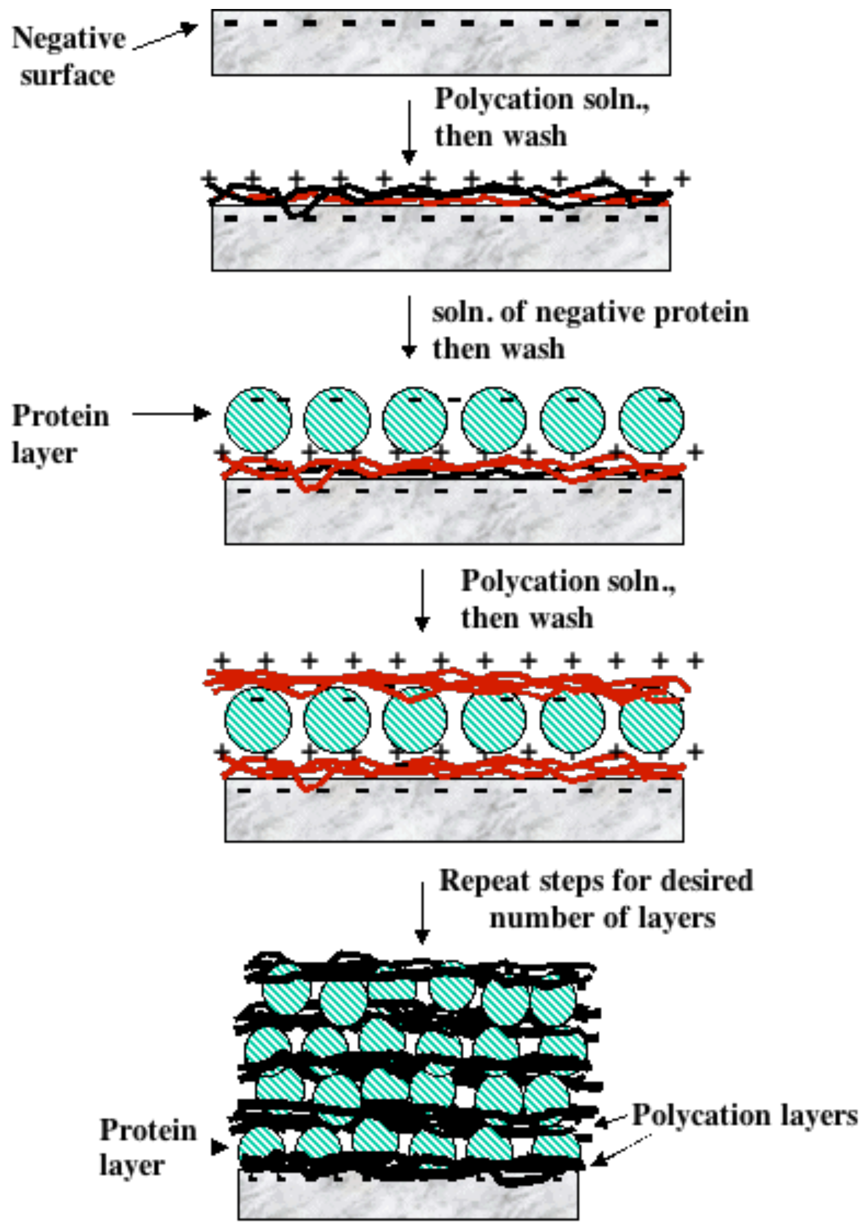
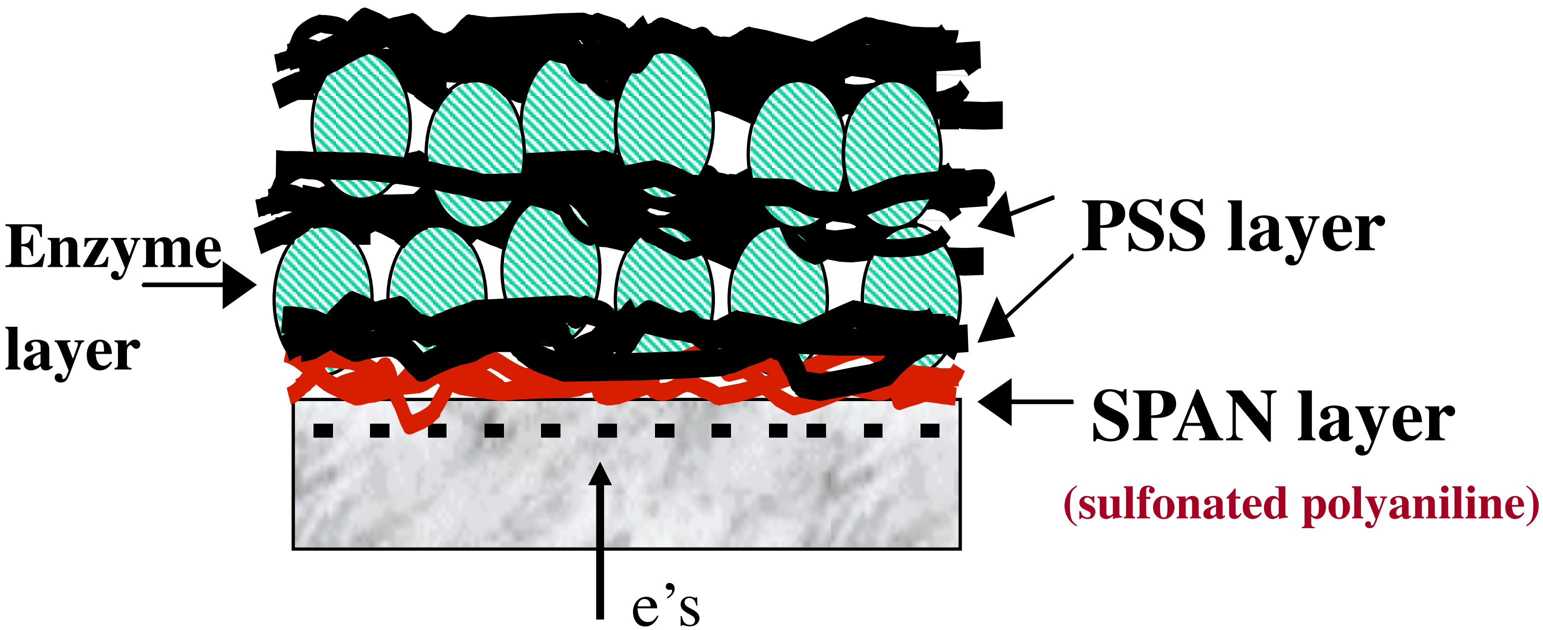


Figure 19

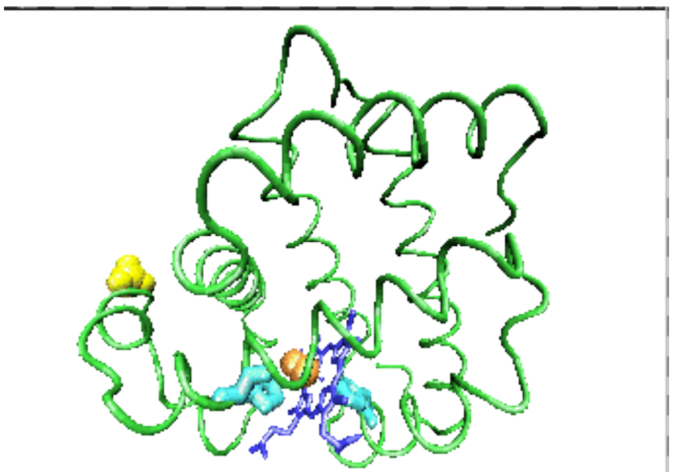
Detection of hydrogen peroxide

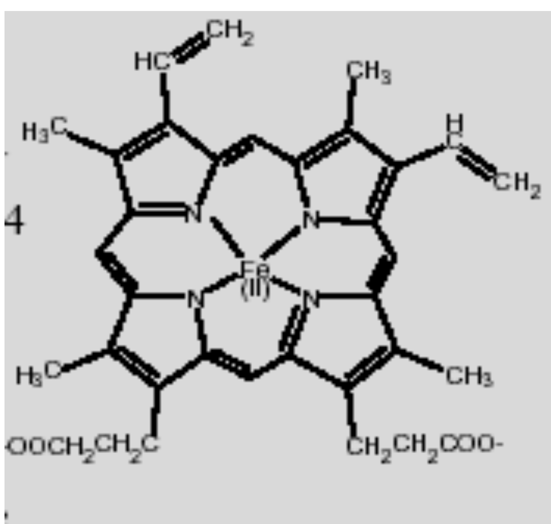
Conductive polymers efficiently wire peroxidase enzymes to graphite

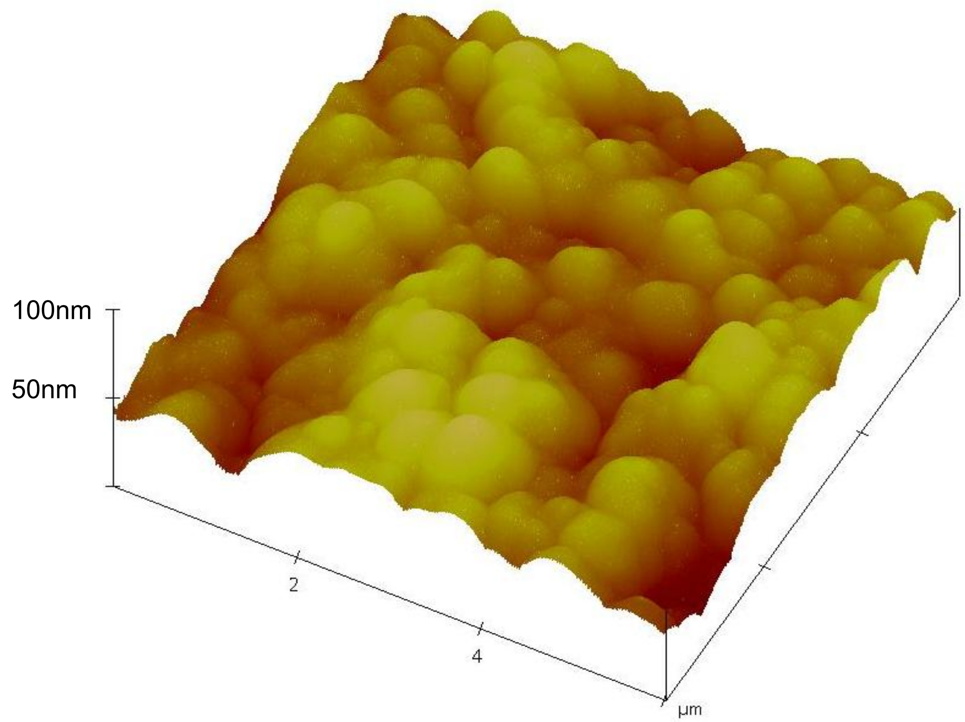


Xin Yu, G. A. Sotzing, F. Papadimitrakopoulos, J. F. Rusling, Highly Efficient Wiring of Enzymes to Electrodes by Ultrathin Conductive Polyion Underlayers: Enhanced Catalytic Response to Hydrogen Peroxide, *Anal. Chem.*, 2003, *75*, 4565-4571.

Horseradish Peroxidase (HRP)

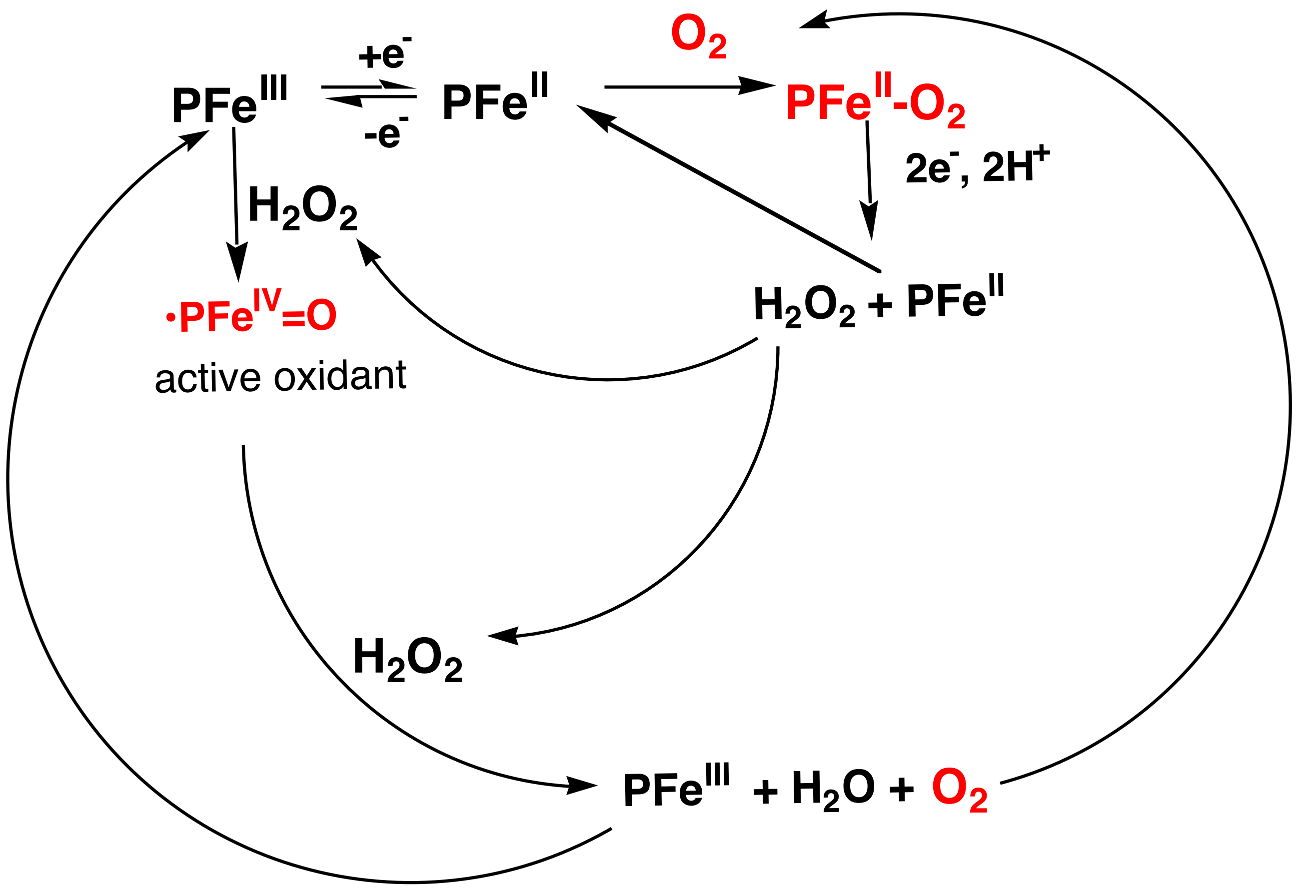






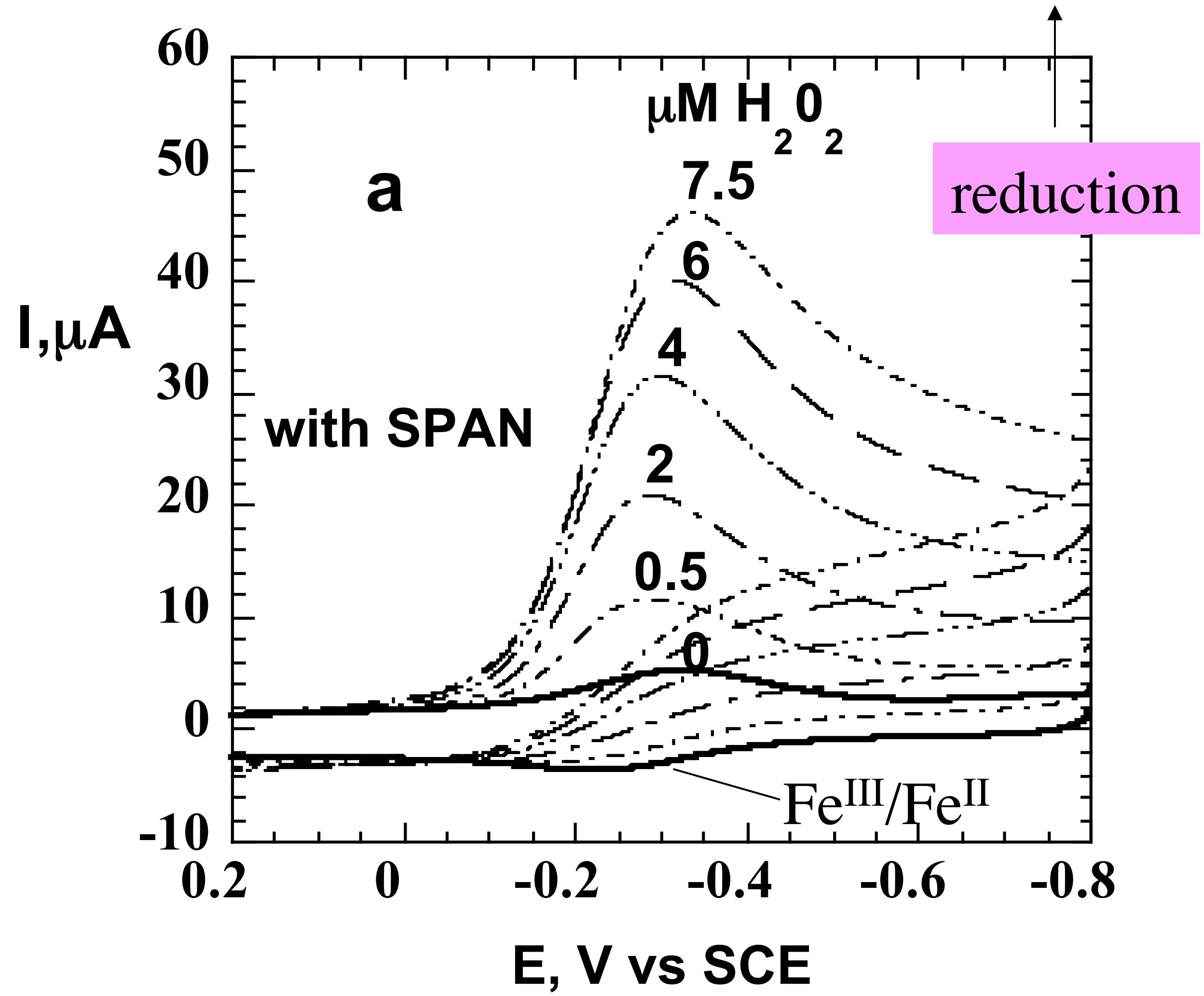
Tapping mode atomic force microscopy (AFM) image of HRP film

Electrochemical Response of Peroxidases

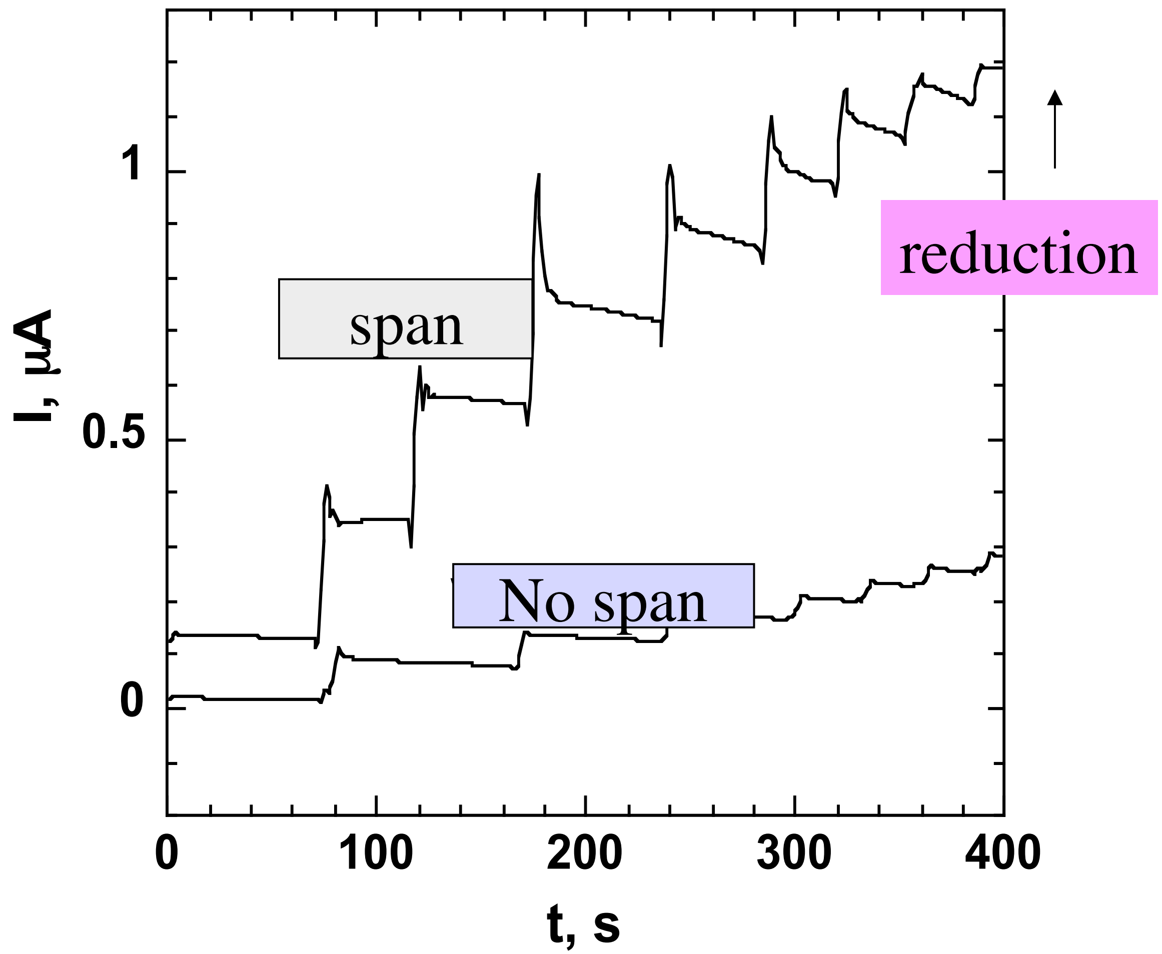


Possible reduced species in red

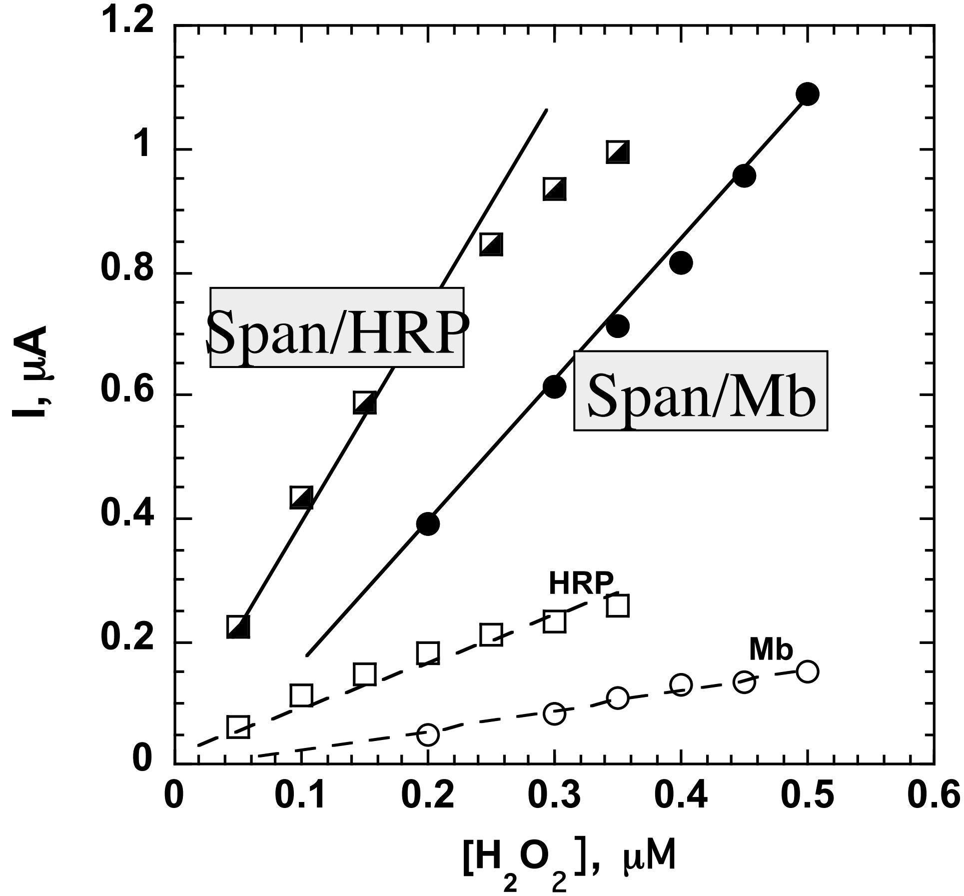
Catalytic reduction of H_2O_2 by peroxidase films
Catalytic cycles increase current



Rotating electrode amperometry at 0 V
HRP, 50 nmol H_2O_2 additions



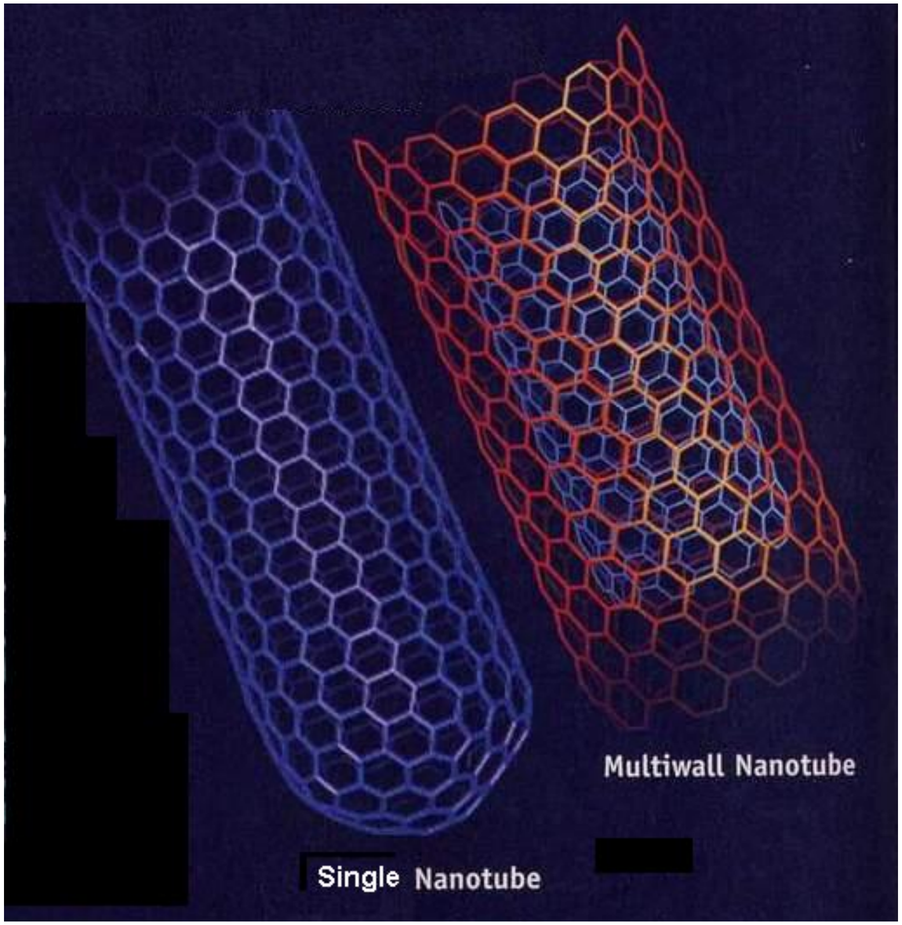
Rotating electrode amperometry at 0 V



Sensitivity much higher with conductive polymer (SPAN); Electrically wires all the protein to electrode

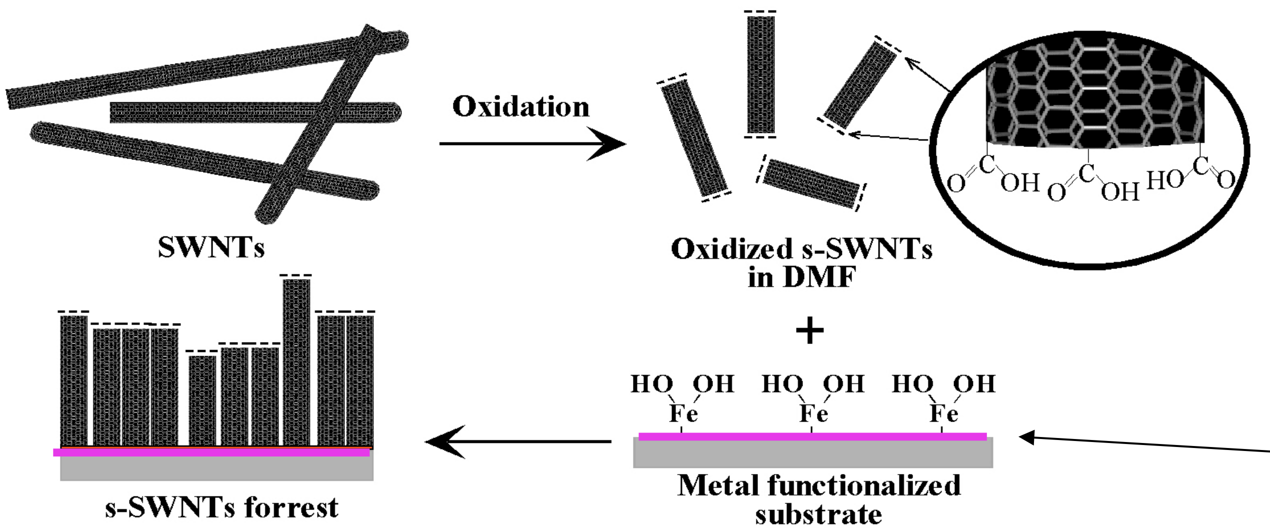
Carbon Nanotubes

- Single walled (1.4 nm o.d.) and multi-walled
- Highly conductive, flexible, strong, patternable
- Commercially Available



Single-Walled Carbon Nanotube Forests: Antigen-Antibody Sensing

~1.4 nm diameter, high conductivity



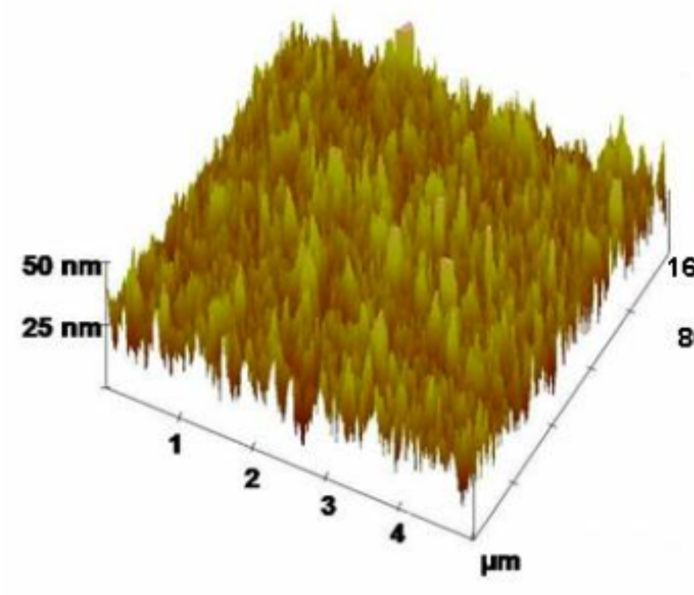
SPAN or Nafion

Chattopadhyay, Galeska, Papadimitrakopoulos, *J. Am. Chem. Soc.* 2001, *123*, 9451.

End COOH groups allow chemical attachment to proteins (antibodies)

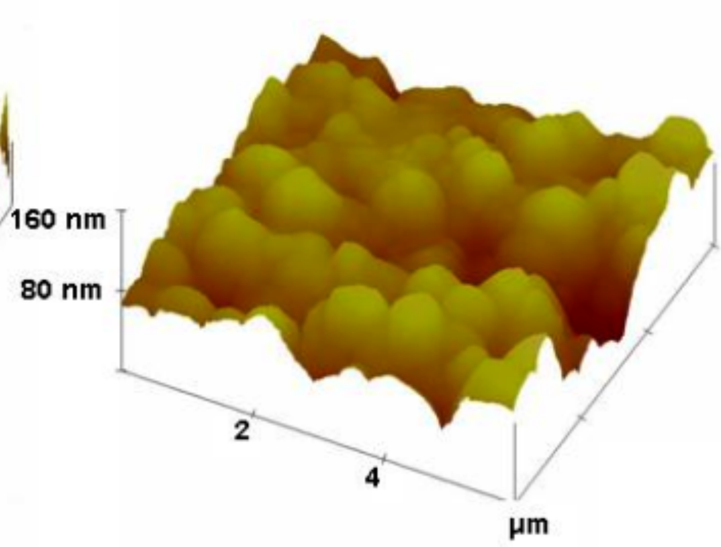
High conductivity to conduct signal (e's) from enzyme label to meas. circuit

AFM of SWNT forest with and without anti-HSA attached



(a)

SWNT forest on Si wafer

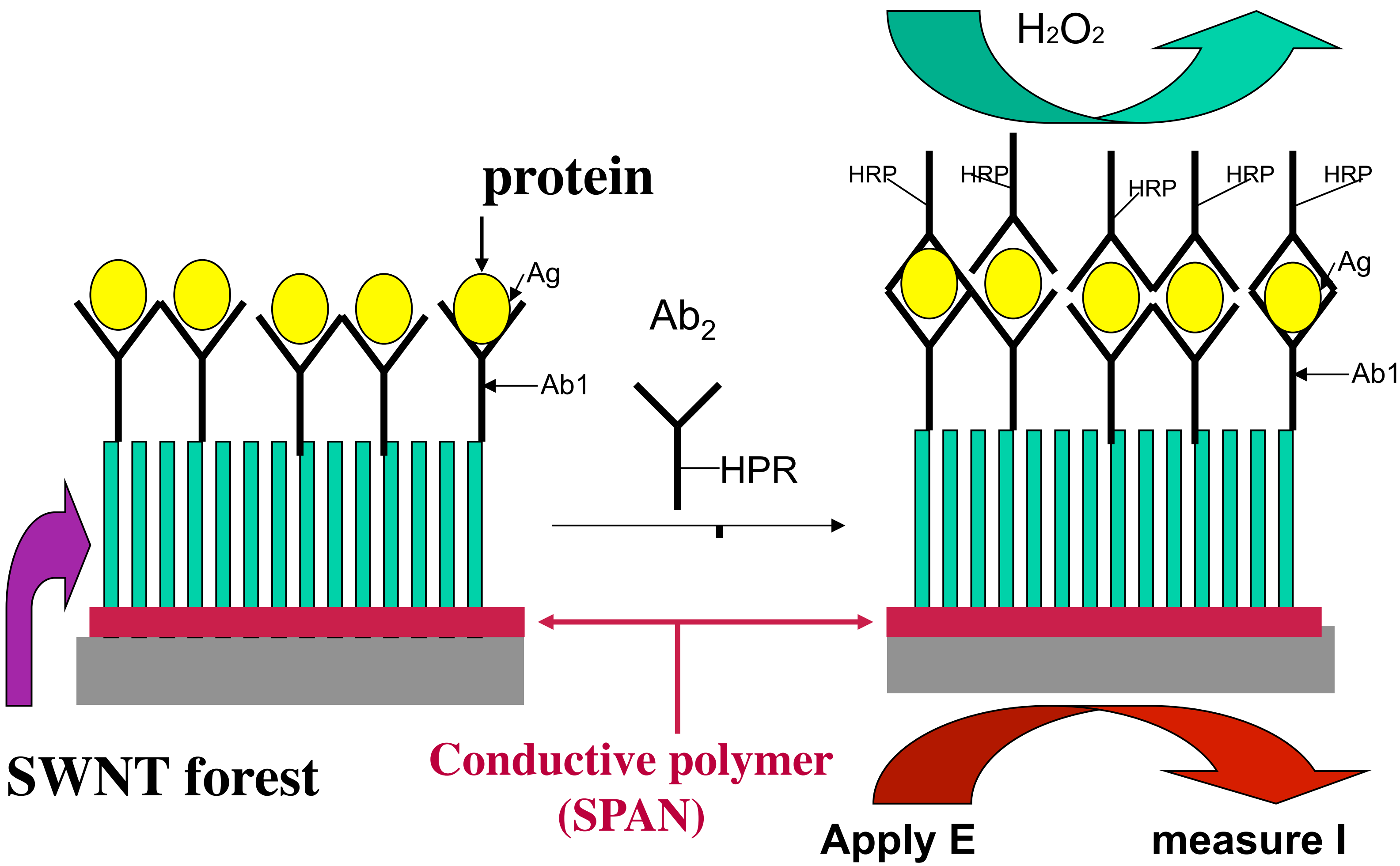


(b)

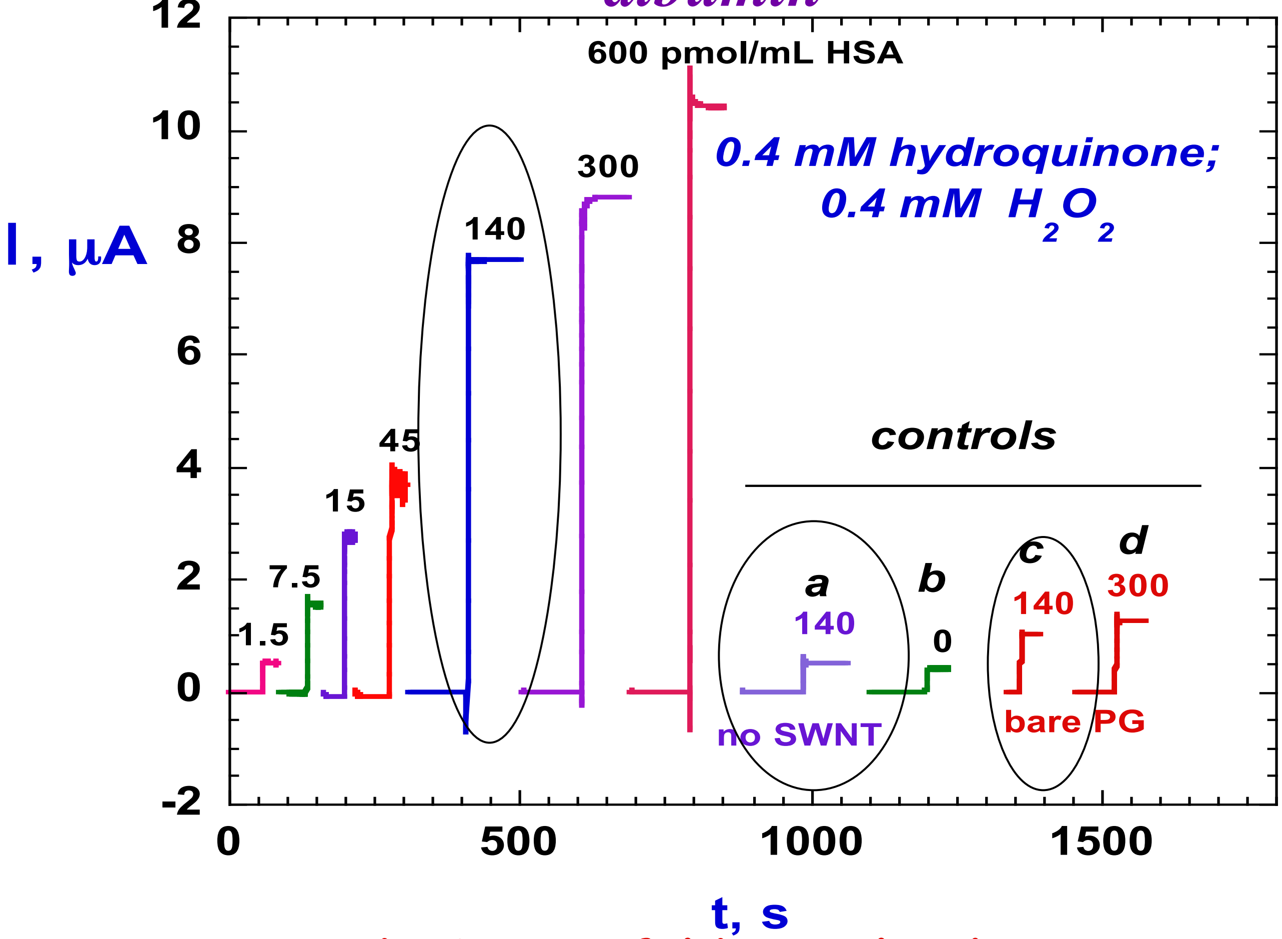
SWNT forest with anti-human serum albumin (HSA) attached by amide links

- **Also linked enzymes to SWNT forests**: X. Yu, D. Chattopadhyay, I. Galeska, F. Papadimitrakopoulos, and J. F. Rusling, "Peroxidase activity of enzymes bound to the ends of single-wall carbon nanotubeforest electrodes", *Electrochem. Commun.*, 2003, *5*, 408-411.

Sandwich Electrochemical Immunosensor Proteins



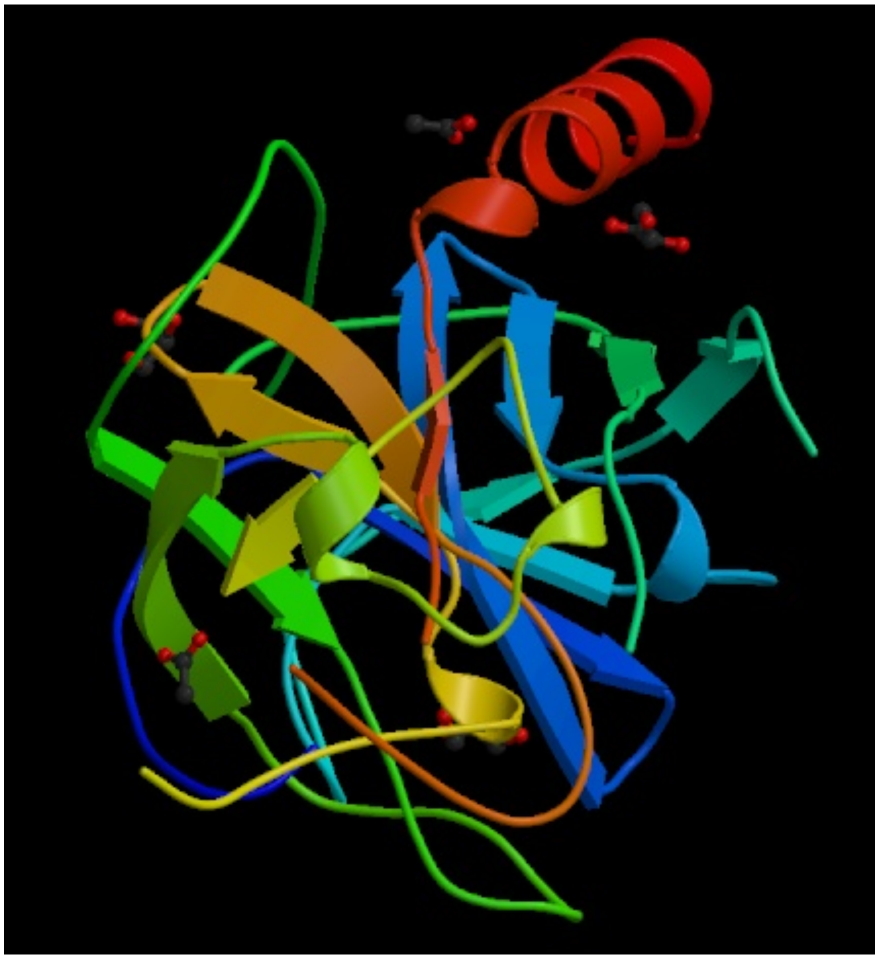
Amperometry Detection of Human Serum albumin



- **SWNTs provide 10-20 fold signal enhancement; high surf. area**
- **Nanotubes aged in DMF ☛ fewer defects ☛ denser forests**

. Xin Yu, Sang Nyon Kim, Fotios Papadimitrakopoulos and James F. Rusling, "Protein Immunosensor Using Single-Wall Carbon Nanotube Forests with Electrochemical Detection of Enzyme Labels", *Molecular Biosystems*, **2005**, *1*, 70-78.

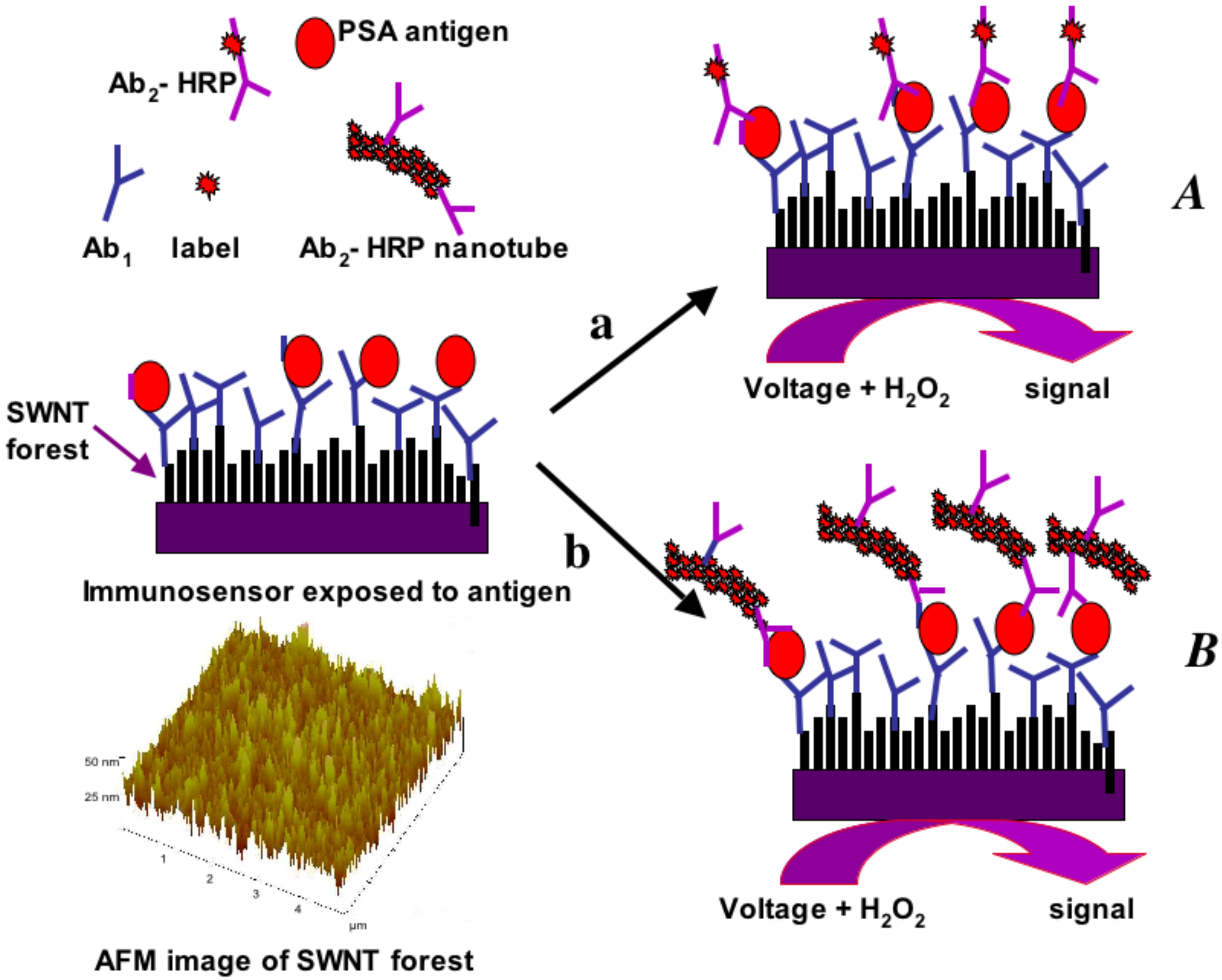
Initial Target: Prostate Specific Antigen



Adapted From Brookhaven Protein Databank

- PSA - Single chain glycoprotein , MW 33 kDa
- Sensitive, specific biomarker for detection of prostate cancer up to 5 years before clinical signs of disease
- Detection of PSA in serum: clinical method for detection of prostate cancer
- Led to less invasive treatment protocols, avoid surgery

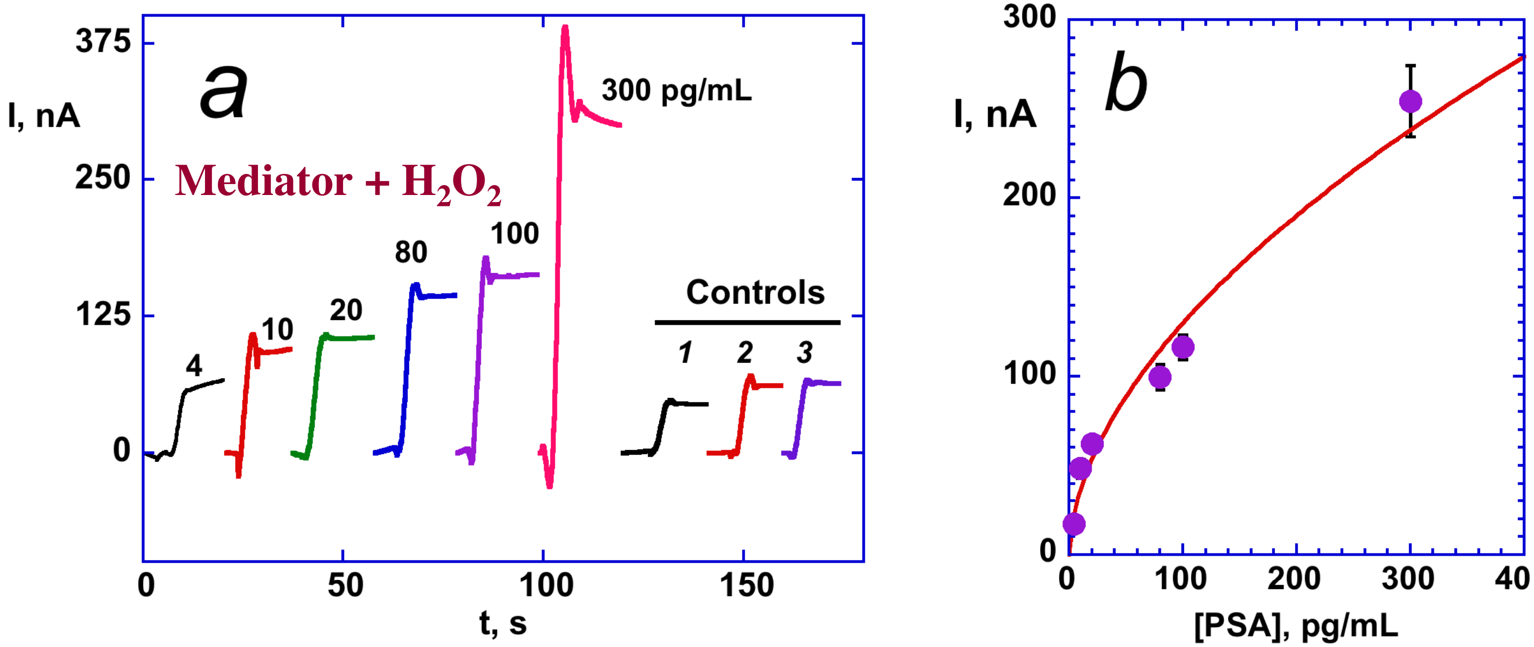
Nanotube Strategies for PSA detection



~170 labels per PSA

Using HRP-Ab_2-nanotube

Amperometric response at –0.3 V and 3000 rpm for SWNT immunosensors incubated with PSA in 10 μL undiluted newborn calf serum for 1.25 hr using the Ab_2-CNT-HRP bioconjugate



Washing with 2% BSA/0.05% Tween 20 to control non-specific binding

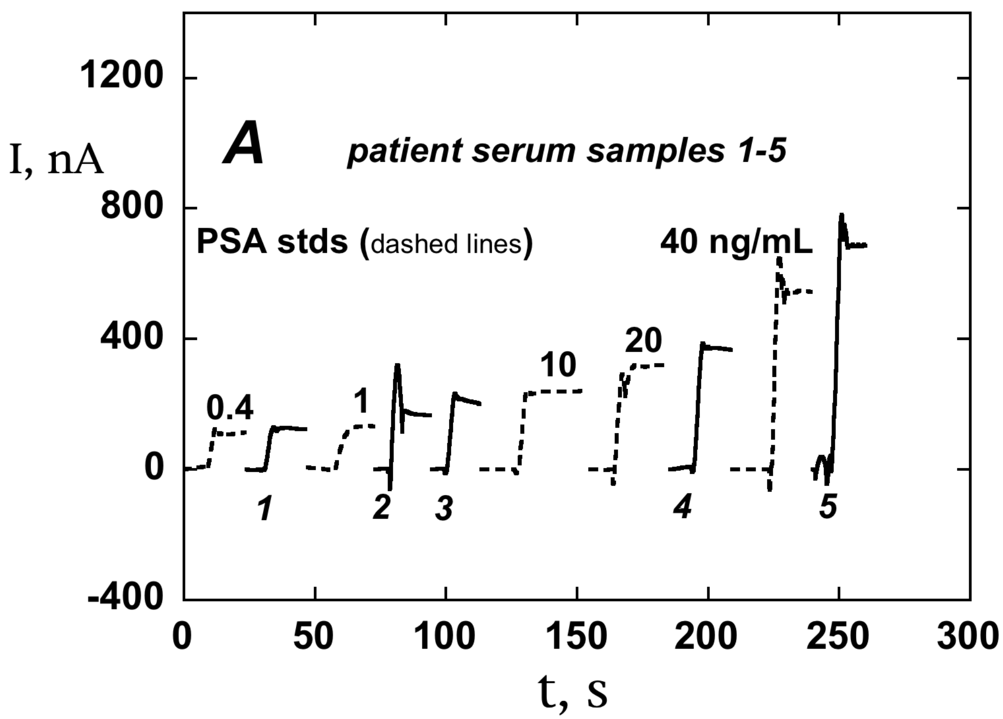
<u>LOD - 4 pg/mL</u>; 100-fold enhancement over HRP-Ab_2

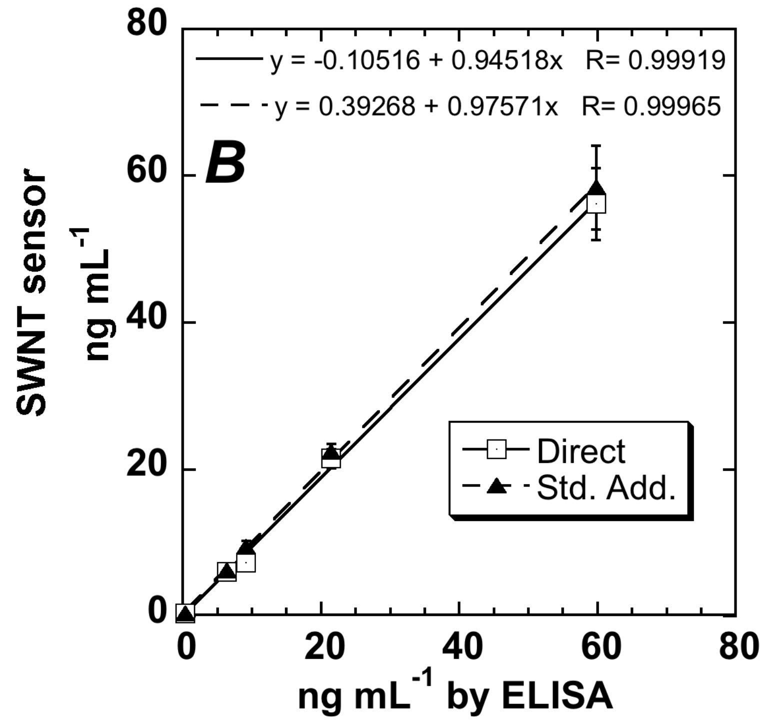
Xin Yu, Bernard Munge, Vyomesh Patel, Gary Jensen, Ashwin Bhirde, Joseph D. Gong, Sang-Nyon Kim, John Gillespie, J. Silvio Gutkind, Fotios Papadimitrakopoulos and James F. Rusling, "Carbon Nanotube Amplification Strategies for Highly Sensitive Immunosensing of Cancer Biomarkers in Serum and Tissue", *J. Am. Chem. Soc.*, **2006**, *128,* 11199-11205.

Accurate results obtained for cancer patient serum

Amperometric current at –0.3 V and 3000 rpm for human serum samples and PSA standards in calf serum

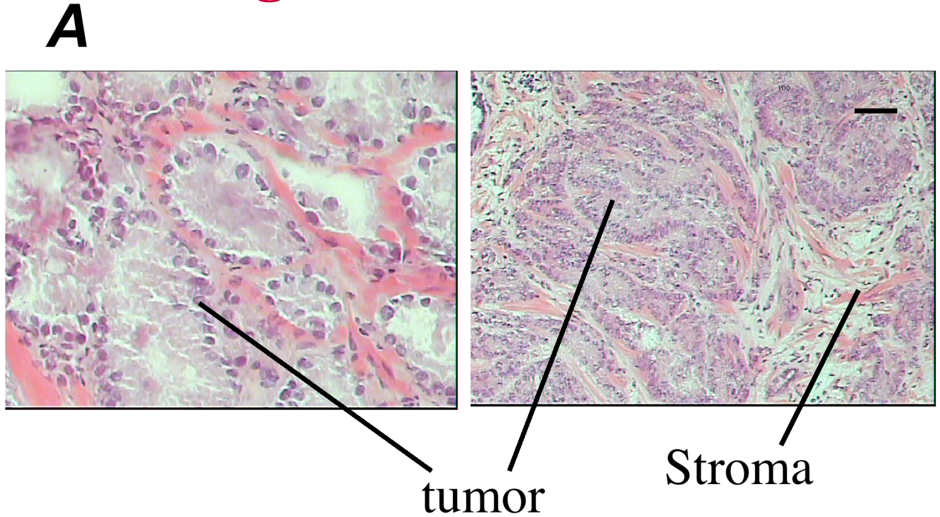
Using conventional HRP-Ab_2

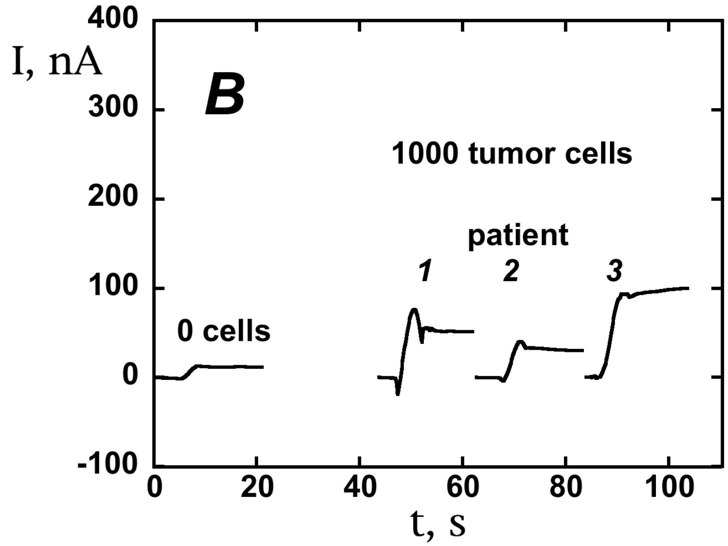




Good correlation with ELISA!

PSA in prostate tissue samples indistinguishable by the best staining methods





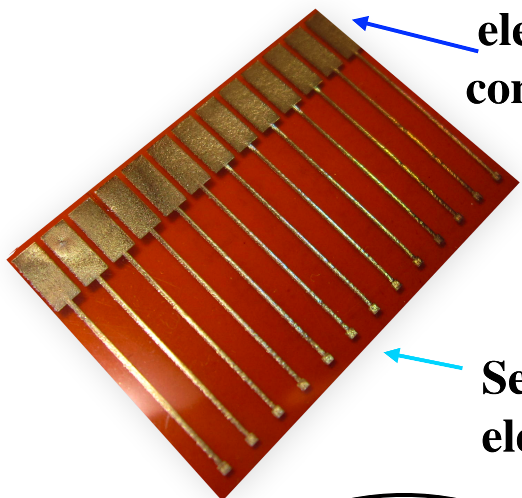
cell lysates obtained after laser microdissection

Using multiply labeled Ab2-nanotube-HRP

4 pg/mL DL (100 x better)

1 **- 90 attogram (ag) PSA/cell, sample *2* - 51 ag PSA/cell, and sample *3* - 830 ag PSA/cell.**

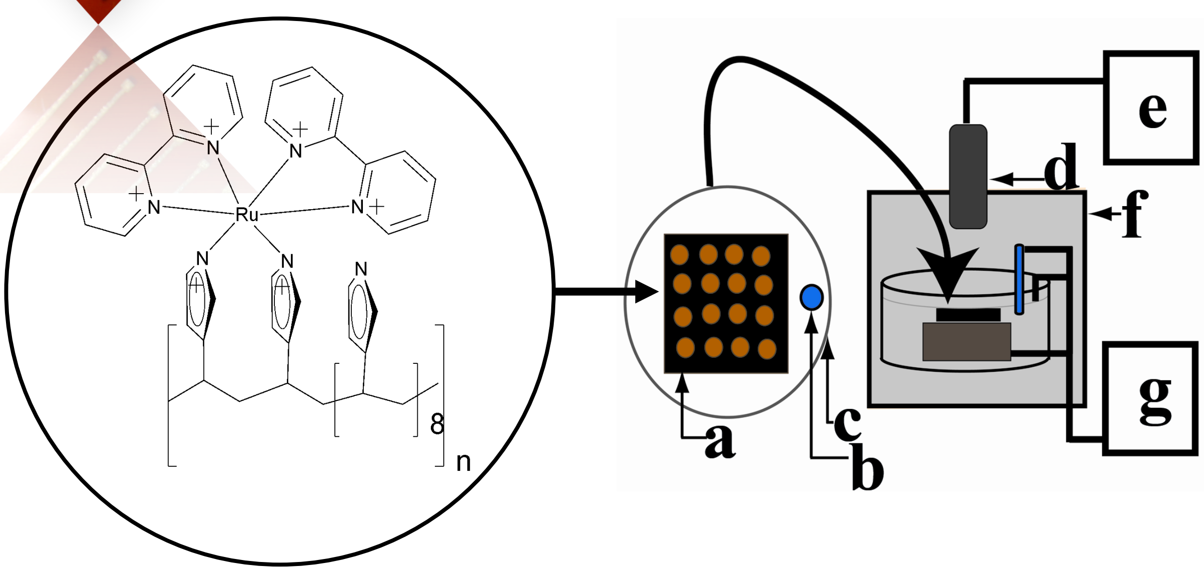
Future - arrays to detect many biomolecules at once



electrical contacts

Sensing electrodes

8-electrode sensor array



An optical-electrochemical array

Polymer gives off light when electro-oxidized with proper co-reactant, e.g. DNA

Biosensors

- **Future of medical diagnostics; use by patients or in doctors offices and clinics**
- **Other important applications: cancer biomarkers, DNA, peroxide, etc.**
- **Method of choice for blood glucose in diabetics**
- **Rapid diagnostics may lead to more timely and effective treatment**